

THE CHEMICAL CONSTITUENTS' OF THE LEAVE ESSENTIAL OIL OF ALTERNANTHERA PUNGENS (KUNTH)

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ABSTRACT

Alternanthera pungens Kunth commonly called khaki weed is from the family Amaranthaceae. It is a herbaceous perennial plant that has stems prostrate, rarely rising and about 10-50 cm long. The work was carried out due to the scarcity of information on the volatile constituents from the plant leaves despite works on the flower and other parts. The extraction of the essential oils from the dried leaves was carried out by the hydro distillation method using an all-glass Clevenger apparatus. The extracted oils were then analyzed using gas chromatography-mass spectrometry (GC-MS). A total of twelve constituents' representing 93.39% of *A. Pungens* oil with a yield of 0.4% (v/w) was obtained. The analysis of the GC-MS results of the leaf oil showed that it was dominated by β -ionone (42.18%) and hexahydrofarnesyl acetone (15.53%), others in trace amounts include; methyl palmitate (6.13%), 1-octadecyne (4.72%), undecane (3.73%), para-mentha-1, 3, 8-triene (3.65%), isophytol (3.21%), d-cadinene (3.06%), 1, 2-dimethyl cyclooctene (3.05%), para-cymene (2.96%), phytol (2.67%) and neophytadiene (2.50%). The common classes of compounds present in the leaves oil are aceto monocyclic monoterpene (42.18%), sesquiterpenoids (18.59%), hydrocarbons (11.50%), diterpenoids (8.38%), monoterpenes (6.61%) and fatty acids (6.13%). The constituents and the compositional pattern of essential oil identified from the leaves of *Alternanthera pungens* grown in Nigeria differ quantitatively and qualitatively from previously reported member of the genus and the presence of sesquiterpenoid as one of the major components of the oils justify the traditional use of the plants in treating pains, headaches and inflammations.

Keywords: *Alternanthera pungens*, Essential oil, β -ionone and Hexahydrofarnesyl acetone.

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INTRODUCTION

The genus *Alternanthera* contains medicinal plants that possess volatile constituents, essential amino acids, flavonoids, glycosides and steroids (Khatun et al. 2012). *Alternanthera pungens* Kunth commonly known as khaki weed in English and 'dágúnró' in Yoruba Southwest Nigeria, belongs to the Amaranthaceae family. It grows widely in tropical and subtropical regions throughout the world (Burkill, 1985). It is an herbaceous perennial plant that has stems prostrate, rarely rising and about 10-50 cm long (Marchant et al., 1987; Naidu, 2012; Hossain et al., 2018). *Alternanthera pungens* is reported to be used traditionally for the treatment of naso-pharyngeal infections, diarrhea, dysentery, vermifuges, dropsy, swellings, oedema, gout and venereal diseases (Zongo et al., 2011; Hundiwalé et al., 2012). It is also used as a painkiller, for treating stomachache, swelling and for lactation stimulus in

veterinary (Burkill, 1985; Mourya et al., 2018).

The plant has been shown to exhibit antioxidant, antimicrobial, antidiabetic, antidiarrhea, spasmogenic, diuretic, and anti-HIV properties (Petruşan and Seetharaman, 2005). The analgesic potential of *Alternanthera pungens* ethanolic extract leaves investigated revealed an improved analgesic effect compared to the standard used. The analgesic effect was attributed to the presence of phytochemicals like flavonoids, tannins and phenolic compounds (Hossain et al., 2018). Furthermore, phytochemicals such as; saponins, alkaloids, steroids, triterpenoids, leucoanthocyanidins, β -spinasterol, saponin sheteroside of oleanolic acid, glycosides, flavonoid stannins, carbohydrate, β -carotene, lutein, violaxanthin, zeaxanthin and choline has been previously described from this plant extracts (Gupta and Saxena, 1987; Mourya et al., 2018).

Occasionally, other genus like *A. sessilis*, *A. philoxeroides*, *A. bettizicikiana* and *A. brasiliana* (flowers, leaves, stems and roots extracts) have been reported to contain similar or related, but not necessarily the same chemical constituents in different percentages. Some of them includes; phaeophytina, β -sitosterol, 3β -hydroxystigmast-5-en-7-one, 24-methylene cycloartanol, cycloeucaleanol, phytol, β -carotene, α -spinasterol, campesterol, triterpenoids, phytosterols, glycosides, alkaloids, flavonoids, steroids, phenolic compounds, carbohydrate, amino acids, proteins, ionone derivatives, saponins, lipids and tannins (De Sooz et al., 1998; Ragasa et al., 2002; Fang et al., 2006; Aguirre et al., 2013; Walter et al., 2014; Vidhya et al., 2015; Devi et al., 2015; Pulipati et al., 2016). The chemical compositions of essential oils (oxygenated sesquiterpenes, diterpenes, ketones, fatty acid and esters) extracted from the flower and leaf of *Alternanthera sessilis* by hydro distillation showed remarkable difference in constituents and percentage composition as well as difference in the antioxidant effect on 1-diphenyl-2-picrylhydrazyl (DPPH) radical scavenging capacities. The major chemical composition of leaves oil are; 1, 1, 1, 5, 5, 5-hexamethyl-3, 3-bis [(trimethylsilyl) oxy] trisiloxane (15.43%) which is an oxygenated diterpenes followed by S, S-dioxide trans-2-methyl-4-N-pentylthiane (11.27%), didodecylphthalate (10.62%) and tetrahydro-2, 5-dimethoxy furan (10.01%) while the prominent ones from flower oil are; 1, 1, 1, 5, 5, 5-hexamethyl-3, 3-bis [(trimethylsilyl) oxy] trisiloxane (17.76%), trans-4-ethyl-5-octyl-2, 2-bis (trifluoromethyl)-1, 3-dioxolane (11.12%) and tetrahydro-2, 5-dimethoxy furan (9.10%). (Khan et al., 2016). Similarly, *A. triandra* seed oil reported a moderate source of ricinoleic acid (22.1%), myristic acid (3.9%), palmitic acid (16.9%), stearic acid (5.90%), oleic acid (26.0%) and linoleic acid (25.2%) (Hosamani et al., 2004). Although, the essential oil of *Alternanthera pungens* flower has been analyzed with the following phytochemicals identified; α -pinene (7.40%), β -pinene (6.42%), camphene (4.21%), myrcene (3.61%), p-cymene (4.29%) limonene (3.52%), β -ocimene (2.35%), 1,8-cineole (6.28%), α -thujone (3.62%), α -borneol (4.46%), α -curcumene (2.36%), camphor (5.52%), linalool (6.29%), geraniol (7.42%), α -terpineol (3.82%), elemol acetate (6.14%), eudesmol (5.38%),

azulene (3.16%) and three unidentified compounds (Gupta and Saxena, 1987). However, there is paucity of information on the essential oil of the leaves. This research is therefore focused on the extraction and determination of the volatile constituents from the leaves of *Alternanthera pungens*.

Materials and Methods

Plant materials

Fresh leaves of *Alternanthera pungens* were obtained from a farm land within Olabisi Onabanjo University, Ago Iwoye, Ogun State, Nigeria. The sample was identified and authenticated at the herbarium of the Forestry Research Institute of Nigeria (FRIN) Ibadan by Mr. A. O. Adeyemo and a voucher specimen with herbarium number FHI-110461 was deposited. The leaves were air dried at room temperature and pulverised before further study.

Extraction of the leaf oil

The air-dried pulverised leaves of *Alternanthera pungens* (500g) was subjected to hydro distillation in a Clevenger-type glass apparatus for 3 h in accordance with established procedure (Ogundajo et al., 2016). The oils collected in hexane was preserved in an amber bottle and stored in a refrigerator for further analysis.

Gas Chromatography-Mass Spectrometry (GC/MS) of the leaf oil.

The GC-MS analysis of the essential oil was carried out on an Agilent model 7890 A gas chromatograph equipped with a FID and fitted with a fused silica capillary HP-5MS column (30 m x 0.32 mm id, film thickness 0.25 μ m). The oven temperature was programmed from 80-240°C at the rate of 80°C/min. The ion source was put at 240°C while electron ionisation was set at 70eV. Helium was used as the carrier gas at a flow rate of 2 mL/min. Scanning range was 35-425 amu. Diluted oil in n-hexane (1.0 μ L) was injected into the GC/MS.

Identification of the Leaf Essential Oil

The identification of the constituents was performed on the basis of retention indices (RI) determined by co-injection with reference to a homologous series of n-alkanes, under identical experimental conditions. Further identification of the components was done by comparison of their



mass spectra with those from NIST (Data base 69) and Wiley 9th Version and the home-made MS library composed of pure substances and components of known essential oils, as well as by comparing their retention indices with literature values (Adams, 2007; Aboaba *et al.*, 2010).

RESULTS

The chemical constituents identified as revealed by GC-MS results as shows in (Fig. 1.0). Aceto monocyclic monoterpenoid (42.18%), sesquiterpenoids (18.59%), hydrocarbons (11.50%), diterpenoids (8.38%), monoterpenes (6.61%) and fatty acids (6.13%). However, sesterterpenes compounds were conspicuously

absent in the essential oil. The yield of essential oil was 0.4% (v/w) calculated on a dry weight basis. The oil sample was cloudy white in colour with a strong herbal odour. A total of twelve constituents representing 93.39% of *A. pungens* oil were identified. In addition, out of the constituents identified, β -Ionone has the highest percentage (42.18%), followed by hexahydrofarnesyl acetone (15.53%), others include; methyl palmitate (6.13%), 1-octadecyne (4.72%), undecane (3.73%), para-mentha-1, 3, 8-triene (3.65%), isophytol (3.21%), α -cadinene (3.06%), 1, 2-dimethyl cyclooctene (3.05%), para-cymene (2.96%), phytol (2.67%) and neophytadiene (2.50%) with the lowest percentage (Table 1.0).

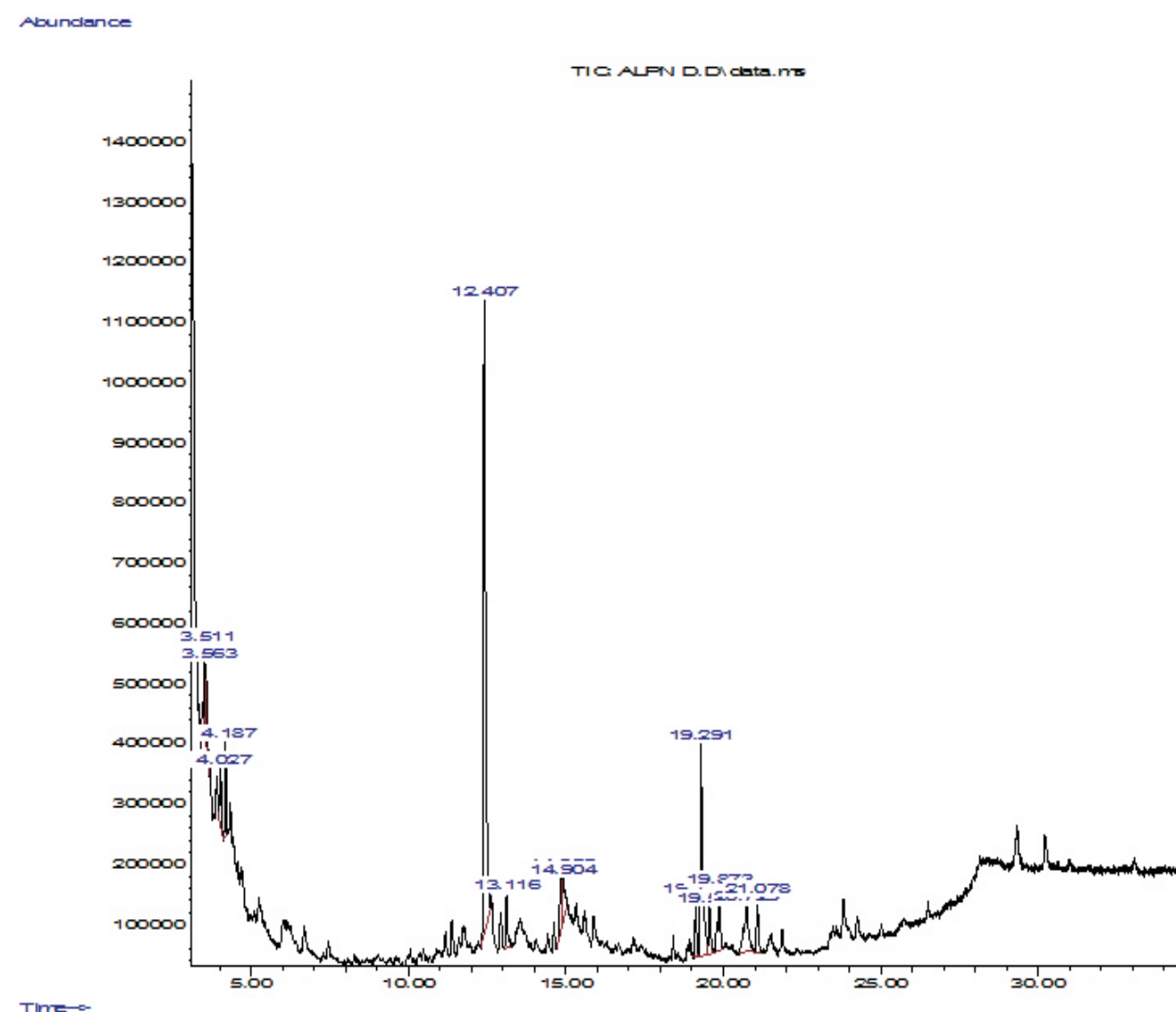


Fig. 1.0: Total Ion Chromatogram of leaf essential oil of *A. pungens*

Table 1.0: The chemical composition of the leaf essential oil of *A. pungens*

S/N	Compounds (common names)	Retention index cal	Retention index lit.	Retention time	% Composition	Molecular formulae
1	1, 2-Dimethyl cyclooctene	986	990	3.511	3.05	C ₁₀ H ₁₈
2	<i>p</i> -Mentha-1,3,8-triene	1000	1110	3.562	3.65	C ₁₀ H ₁₄
3	<i>p</i> -Cymene	1074	1073	4.025	2.96	C ₁₀ H ₁₄
4	Undecane	1115	1100	4.186	3.73	C ₁₁ H ₂₄
5	β -Ionone	1468	1461	12.408	42.18	C ₁₃ H ₂₀ O
6	δ -Cadinene	1374	1530	13.118	3.06	C ₁₅ H ₂₄
7	Neophytadiene	1810	1827	19.126	2.50	C ₂₀ H ₃₈
8	Hexahydrofarnesyl acetone	1954	1858	19.292	15.53	C ₁₈ H ₃₆ O
9	Phytol	2105	2111	19.566	2.67	C ₂₀ H ₄₀ O
10	1-Octadecyne	1920	1238	19.875	4.72	C ₁₈ H ₃₄
11	Methyl palmitate	1942	1928	20.728	6.13	C ₁₇ H ₃₄ O ₂
12	Isophytol	1899	1920	21.077	3.21	C ₂₀ H ₄₀ O

DISCUSSIONS AND CONCLUSION

The quantitative amount of β -ionone in this oil is noteworthy (42.18%), since it has not been previously reported to be a major compound of *A. pungens*. β -Ionone is one of essential constituents in most aromatic oils used in the production of perfumes due to its strong pleasant odour (Cseke *et al.*, 2007). Furthermore, β -ionone has been reported to be fragrance ingredient in perfumery, cosmetics, personal care products, household cleaners and detergents. It is also a flavouring agent in beverages, baked goods, and candies. β -ionone is of use, not only in perfumery, but also as a key intermediate in the synthesis of Vitamins A, E and K (Lalko *et al.*, 2007). Not surprisingly, the steam distillation of the flower essential oil (0.6%) of *A. pungens* has a very different composition compared to the leaf oils. These includes; α -pinene (7.40%), β -pinene (6.42%), camphene (4.21%), myrcene (3.61%), para-cymene (4.29%), limonene (3.52%), β -ocimene (2.35%), cineole (6.28%) among others (Gupta and Saxena, 1987). Previous report on essential oils of another specie; *A. sessilis* from Malaysia showed the major components of leaves as 1,1,1,5,5,5-hexamethyl-3,3-bis [(trimethylsilyl) oxy] trisiloxane (15.43%), S, S - dioxide trans - 2 - methyl - 4 - N - pentyl

thiane (11.27%), didodecylphthalate (10.62%) and tetrahydro-2, 5-dimethoxy furan (10.01%), unlike the current work that was majorly dominated by β -ionone (42.18%) and hexahydrofarnesyl acetone (15.53%). The other characteristics compounds previously reported from *A. sessilis* were; octyl ester-2-propenoic acid (9.32%), 1-fluorododecane (6.60%), 1-chlorooctadecane (3.99%), methyl-2-O-methyl- α -D-xylofuranoside (3.13%), (Z)-7-tetradecene (3.12%), trimethyl (4-terbutylphenoxy) silane (2.98%), 1,1'-oxybis-heptane (2.41%), cis-4-penty 1-5-propyl-2, 2-bis (trifluoromethyl)-1, 3-dioxane (2.29%) and 9-heptadecanone (2.06%) (Khan *et al.*, 2016).

The chemical compositions of essential oil from the leaves of *A. pungens* grown in Nigeria are being reported for the first time based on available information. The analysis of the extracted essential oil from *A. pungens* showed the presence of aceto monocyclic monoterpenoids (42.18%), sesquiterpenoids (21.41%) and hydrocarbons (11.50%) as the major predominant classes of compounds. Oxygenated diterpenes (11.27%) was reported as the most prominent from the leaf oil of *A. sessilis* (Khan *et al.*, 2016) as compared to aceto monocyclic monoterpenoids (42.18%) which is the major class in the present study. It was observed that constituents and the compositional pattern of

essential oil from the leaves of *Alternanthera pungens* grown in Nigeria differ quantitatively and qualitatively from previously reported member of the genus (Khan *et al.*, 2016). This may be attributable to factors such as the ecological and climatic conditions as well nature and age of the plant, period of collection, handling procedures and other related factors known to induce variation in essential oil composition extracted from plants (Gurusaravanan *et al.*, 2010; Verma *et al.*, 2013; Inikpi *et al.*, 2014; Rehman *et al.*, 2016; Ibanez and Blazquez, 2019). Conclusively, the chemical compositions of essential oil from the leaves of *A. pungens* grown in Nigeria are being reported for the first time based on available information. The presence of sesquiterpenoid as one of the major classes of compounds of the oils justifies the traditional use of the plants in treating pains, headaches and inflammations. Also, the identified chemical constituents of the plant are an addition to the chemical data base for the plant.

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References

- Aboaba, S. A., Aiyelaagbe, O. O. and Ekundayo, O. (2010). Chemical Composition, Toxicity and Larvicidal Activity of the Essential Oil from the Whole Plant of *Acalypha segetalis* from South-West Nigeria. *Natural Product Communications*, 5(3): 481-483.
- Adams, R. P. (2007). Identification of Essential Oil Components by Gas Chromatography/Mass Spectrometry (4th edition), Allured Publishing Corp., Carol Stream, IL, USA.
- Aguirre, A., Borneo, R. and Le'on A. E. (2013). Antimicrobial, mechanical and barrier properties of tritcale protein films incorporated with oregano essential oil. *Food Bioscience*, 1: 2-9.
- Burkill, H. M. (1985). The useful plants of west tropical Africa. *Royal Botanic Gardens*, Kew, UK. 3.
- Cseke, L. J., Kaufman, P. B., and Kirakosyan, A. (2007). The biology of essential oils in the pollination of flowers. *Natural Product Communications*, 2(12):1317-1336.
- De Souza, M. M., Ana, P. K., Floriani, E. O. and Cechinel-Filho, V. (1998). Analgesic Properties of a Hydroalcoholic Extract Obtained from *Alternanthera brasiliensis*. *Phytotherapy Research*, 12: 279-281
- Devi, A. D., Singh, T. C., Devi, O. I., Singh, S. S., Singh, A. R. and Singh, E. J. (2015). Phytochemical Analysis of Some Traditional Aromatic Plant Species of Thoubal District, Manipur. *Asian Journal of Pharmaceutical Science and Technology*, 5(1): 50-53.
- Fang, J. B., Duan, H. Q., Zhang, Y. W. And Yoshihisa, T. (2006). Chemical constituents from herb of *Alternanthera philoxeroides*. *China Journal of Chinese Material Medica*, 31(13): 1072-1075.
- Gupta, R. K. and Saxena, V. K. (1987). Volatile constituents from the flowers of *Alternanthera pungens* HBK (Amaranthaceae). *Indian Perfume*, 31(4): 366-369.
- Gurusaravanan, P., Pandiyarajan, V. and Jayabalan, N. (2010). Effect of the seaweed liquid fertilizer on growth and productivity of *Vignaradiata* (L.) Wilczek. *Green Farm*, 1(2): 138-140.
- Hosamani, K. M., Ganjihal, S. S. and Chavadi, D. V. (2004). *Alternanthera triandra* seed oil: A moderate source of ricinoleic acid and its possible industrial utilization. *Industrial Crops and Products*, 19: 133-136.
- Hossain M. D. S., Rana, M. D. S., Rajibul Islam, Mahmudul Islam, A.F. M. (2018). Phenolic content analysis and evaluation of antinociceptive, antioxidant, anti-inflammatory potential of *Alternanthera pungens* KUNTH. *Journal of Bangladesh Academy of Sciences*, 42(2): 129-136
- Hundiware, J. C., Patil, A. V., Kulkarni, M. V., Patil, D.A. and Mali, R. G. (2012). A current update on phytopharmacology of the genus *Alternanthera*. *Journal of Pharmacy Research*, 5(4): 1924-1929.
- Ibanez, M. D. and Blazquez, M. A. (2019). Essential Oils: Quality Indicators of Spices in Supermarkets. Nereis. *Interdisciplinary Ibero-American Journal of Methods, Modelling and Simulation* Marzo, 11: 39-50
- Inikpi, E., Lawal, O. A., Ogunmoye, A. O. and Ogunwande, I. A. (2014). Volatile composition of the floral essential oil of *Hibiscus sabdariffa* L. from Nigeria. *America Journal of Essential Oils and Natural Products*, 2(2): 4-7.
- Joulain, D., Koenig, W. A and Verlag, E.B, Hamburg. 1998 (1999). The Atlas of Spectra Data of Sesquiterpene Hydrocarbons. *Journal of Natural Product*, 62(8): 1212-1213.
- Khan, M. S., Yusufzai, S. K., Kaun, L. P., Shah, M. D. and Idris, R. (2016). Chemical composition and antioxidant activity of essential oil of leaves and flowers of *Alternanthera sessilis* red from Sabah. *Journal of Applied Pharmaceutical Science*, 6(12): 157-161
- Khatun, F., Zaman, F., Mosaib, T., Mostafa, F., Zaman, M., Rehana, F., Nasrin, D., Jamal, F., Nahar, N. and Rahmatullah, M. (2012). Evaluation of antinociceptive and antihyperglycemic activities in methanol extracts of whole plants of *Alternanthera philoxeroides* (Mart.) Griseb. (Amaranthaceae) in mice. *Pakistan Journal of Pharmaceutical Sciences*, 25: 583-587
- Lalko, J., Lapczynski, A., McGinty, D., Bhatia, S., Letizia, C. S. and Api, A. M. (2007) Fragrance material review on trans-beta-ionone. *Food and Chemical Toxicology*, 45(1): 248-50.
- Marchant, N.G., Wheeler, J.R., Rye, B.L., Bennett, E.M., Lander, N.S. and Macfarlane, T.D. (1987). Flora of the Perth Region. (Western Australian Herbarium, Department of Agriculture, Western Australia). p92.
- Mourya, P., Singh, G., Jain, N., Gupta, M. K. (2018). In-vitro studies on inhibition of alpha amylase and alpha glucosidase by plant extracts of *Alternanthera pungens* KUNTH. *Journal of Drug Delivery & Therapeutics*, 8(6-A): 64-68
- Naidu, V. S. G. R. (2012). Hand Book on Weed Identification. *Directorate of Weed Science Research*, Jabalpur, India. p. 354
- Ogundajo, A. L., Nnaemeka, C. O., Olawunmi, R. O. and Ogunwande, I. A. (2016). Chemical Constituents of Essential oil of *Ethretia cymosa* Thonn. *British Journal of Applied science and Technology*, 14(4):1-6
- Petrus, A. J. A. and Seetharaman, T. R. (2005). Antioxidant flavone C-biosides from the aerial parts of *Alternanthera pungens*. *Indian Journal of Pharmaceutical Sciences*, 67(2): 187-193.
- Pulipati, S., Srinivasa, B. P., Sree, N. B., Kumar, U. E., Shaheela, S., Krishna, J. M. and Chakradhar, T. (2016). Phytochemical Analysis and Antimicrobial Investigations of Ethanolic Leaf Extract of *Alternanthera philoxeroides* (MART.) GRISEB. *World Journal of Pharmacy and Pharmaceutical Sciences*, 5(2): 1122-1129.
- Ragasa, C. Y., Tremor, N. and Rideout, J. A. (2002). Ionone derivatives from *Alternanthera sessilis*. *Journal of Asian Natural Products Research*, 4(2): 109-115.
- Rehman, R., Hanif, M. A., Mushtaq, Z. and Al-Sadi, A.M. (2016). Biosynthesis of essential oils in aromatic plants: a review. *Food Reviews International*, 32(2): 117-160.
- Verma, R.S., Padalia, R. C. and Chauhan, A. (2013). Introduction of *Cymbopogon distans* (Nees ex Steud.) Wats to the sub-tropical India: evaluation of essential-oil yield and chemical composition during annual growth. *Industrial Crops and Product*, 49: 858-863.
- Vidhya, T., Suji, T., Dhatchayani, R., Priya, C. L. and Bhaskara-Rao, K. V. (2015). Evaluation of In-vitro Antioxidant, Antimicrobial Activities and GC-MS Analysis of *Alternanthera bettizickiana* Linn. Leaf Extracts. *International Journal of Pharmacognosy and Phytochemical Research*, 7(6): 1072-1079
- Walter, T. M., Merish, S. and Tamizhamuthu, M. (2014). Review of *Alternanthera sessilis* with Reference to Traditional Siddha Medicine. *International Journal of Pharmacognosy and Phytochemical Research*, 6(2): 249-254
- Zongo, C., Savadogo, A., Somda, M. K., Koudou, J. and Traore, A. S. (2011). In vitro evaluation of the antimicrobial and antioxidant properties of extracts from whole plant of *Alternanthera pungens* H.B. & K. and leaves of *Combretum sericeum* G. Don. *International Journal of Phytomedicine*, 3: 182-191